IACUC & LAP

INSTITUTIONAL ANIMAL CARE AND USE COMMITTEE AND THE LABORATORY ANIMAL PROGRAM

MARCH 2024

RELATED WEBSITES

IACUC website—for forms, policies, guidelines, links for Occupational Health & Animal Qualifications info, etc.: www.purdue.edu/animals

Occupational Health Info: https://www.purdue.edu/research/regulatory-affairs/animal-research/occupational-health.php

PERA Project

Many of you may be aware that the IACUC office is actively working on replacing our current on-line protocol system (Coes). The PERA (Purdue Excellence in Research Administration) project includes the IACUC module, as well as other modules used by faculty and staff at Purdue for their research administration needs. The PERA project will utilize Huron Research Systems software to implement these modules.

The timeline for going live with the IACUC module is currently set for late summer/early fall 2024. This timeline will remain fluent based on progress made to integrate from Coeus to the PERA system.

I am currently working on the IACUC module in PERA by populating Specification forms and Bulk Import Sheets with Purdue-specific needs. The major change that animal users will see when PERA is implemented is the IACUC module will go to the use of Smart Forms for the protocol application versus the current Word document format that we currently use in Coeus.

I will continue to keep you updated on this project as it progresses. Please reach out to me if you have any questions/ concerns.

Lisa Snider, IACUC Administrator

AHEF Forms have been updated and will be effective March 1, 2024.

LAP has updated the Animal Health Evaluation Form (AHEF). Please note the change that a response time of "Next Purdue business day by noon" has been added. This response time may be used for mild cases on weekends and holidays, and assumes that the investigative staff will be responsible for checking and responding to LAP by noon of the following business day. Business days are M-F, excluding Purdue holidays or when the campus is closed. This new form will go into effect March 1, 2024. The updated form will come from CMAF staff.

Please contact LAPvet@groups.purdue.edu for any questions.

REMINDER!

Online Form for Self-Reporting of Adverse Events and/or Non-Compliance

The IACUC Committee is pleased to introduce a new online form to report all adverse and non-compliance events. Principal investigators and lab personnel may now notify IACUC of issues by using the online Adverse Event Reporting Form. Completed forms must be sent to the IACUC email address: IACUC@purdue.edu. The link to the new form is: www.purdue.edu/research/oevprp/regulatory-affairs/animal-research/docs/Purdue%20IACUC%20Noncompliance%20-%20Adverse%20Event%20Reporting%20Form.pdf

As a reminder, adverse events are incidents that lead to unanticipated injury or illness, unrelieved pain or distress, or the death of an animal. Non-compliance events occur when an IACUC protocol, policy, guideline, SOP, or procedure is not followed.

Adverse events and non-compliance issues must be promptly reported to the IACUC office.

Assistance Available for Creating and Formatting Search Strings

The IACUC office is offering assistance with creating or formatting search strings that will be listed in the Alternative Search tab in Coeus Lite. If you would like assistance with this, please contact Nancy Mantick, IACUC Protocol Analyst, at mantick@purdue.edu (email is the preferred mode of contact), and provide your keywords for your searches. Making sure your search strings are formatted properly before submitting your new protocol or renewal can help it move more smoothly through the IACUC approval process. Please contact Nancy **PRIOR TO** submitting your protocol application (or triennial renewal for assistance with search strings.

CMAF Per Diem Changes

CMAF Per Diems have been updated on the LAP Website. There is a pull down menu to show future rate increases. Go to https://www.purdue.edu/research/oevprp/regulatory-affairs/animal-research/cmaf/animal-per-diem.php

CMAF Per Diems

All costs are per animal unless noted otherwise. Rates effective on January 1. (*Indicates approved rates. All others are projected at 7% annual increase for grant planning.)

	2023*	2024*	2025	2026	2027	2028
Chinchilla	$2.28	$ 2.39	$ 2.56	$ 2.74	$ 2.93	$ 3.13
Cow (stall)	$15.00	$ 15.75	$ 16.85	$ 18.03	$ 19.29	$ 20.65
Dogs (teaching – research)	$8.50	$ 8.93	$ 9.56	$ 10.22	$ 10.94	$ 11.71
Frogs per tank	$2.42	$ 2.54	$ 2.72	$ 2.91	$ 3.11	$ 3.33
Guinea Pigs	$1.69	$ 1.77	$ 1.89	$ 2.03	$ 2.17	$ 2.32
Horse (stall)	$18.72	$ 19.66	$ 21.04	$ 22.51	$ 24.08	$ 25.77
Mice: Conventional Housing - Per Cage	$1.08	$ 1.13	$ 1.21	$ 1.29	$ 1.38	$ 1.48
Mice: Vent Rack (non-sterile) Per Cage	$0.78	$ 0.82	$ 0.88	$ 0.94	$ 1.00	$ 1.07
Mice: Vent Rack (autoclave in or out) Per Cage	$0.99	$ 1.04	$ 1.11	$ 1.19	$ 1.27	$ 1.36
Mice: Static (autoclave, in or out) Per Cage	$1.35	$ 1.42	$ 1.52	$ 1.63	$ 1.74	$ 1.86
Mice: Vent Rack (autoclave in and out) Per Cage	$1.36	$ 1.43	$ 1.53	$ 1.64	$ 1.75	$ 1.87
Mice: Static (autoclave, in and out) Per Cage	$1.50	$ 1.58	$ 1.69	$ 1.81	$ 1.94	$ 2.07
Poultry (on floor) *	$1.21	$ 1.27	$ 1.36	$ 1.45	$ 1.56	$ 1.66
Poultry (large brooder) * Per brooder	$3.25	$ 3.41	$ 3.65	$ 3.90	$ 4.18	$ 4.47
Poultry (small brooder) per brooder	$4.45	$ 4.67	$ 5.00	$ 5.35	$ 5.72	$ 6.12
Sheep (Pen / Stall)	$6.50	$ 6.83	$ 7.31	$ 7.82	$ 8.37	$ 8.95
Rabbits	$3.64	$ 3.82	$ 4.09	$ 4.37	$ 4.68	$ 5.01
Rats: Conventional Housing	$1.04	$ 1.09	$ 1.17	$ 1.25	$ 1.34	$ 1.43
Rats: Static or Vent (autoclave in or out)	$1.30	$ 1.37	$ 1.47	$ 1.57	$ 1.68	$ 1.80
Rats: Vent Rack (autoclave in and out)	$1.64	$ 1.72	$ 1.84	$ 1.97	$ 2.11	$ 2.25
Swine **	$6.44	$ 6.76	$ 7.23	$ 7.74	$ 8.28	$ 8.86
Swine (pig turn)	$7.25	$ 7.61	$ 8.14	$ 8.71	$ 9.32	$ 9.98
Technical Assistance per hour	$27.29	$ 34.63	$ 37.05	$ 39.65	$ 42.42	$ 45.39
Technical Assistance Overtime per hour	$40.93	$ 51.95	$ 55.59	$ 59.48	$ 63.64	$ 68.10
Stall Set up Fee	$38.00	$ 39.90	$ 42.69	$ 45.68	$ 48.88	$ 52.30

External Rates Availabe On Request

PURDUE UNIVERSITY | Office of Research

IACUC Policy Donation of Rodents for Feed – SOP #502

Purpose: To identify which animals are appropriate for donating to wildlife rehabilitation centers, zoos, etc., which centers are approved for donations, and how to donate to such entities.

Background: The Purdue University IACUC authorizes the donation of rodents as a food source according to the criteria described below. Non-profit organizations with the mission of rehabilitating and releasing injured and orphaned raptors back into the wild may wish to obtain mice and rats previously used for research and/or teaching purposes as a source of food for raptors and other species in its care.

Rodent Donations

In order for an investigator to donate mice or rats as feed, the "Other" box must be selected under the Carcass Disposition section (7.5) on the protocol, it must be specified where the animals are being donated to, and the protocol must be approved. The animals must meet the criteria below to be acceptable for donation. Donations must be delivered to animal facility managers, who will then place the carcasses in the appropriate container in the freezer located in AHF. Donations will be picked up by the accepting entity on a monthly basis, which will be coordinated with the Laboratory Animal Program.

Donation Criteria

- Only wild-type mice or rats may be donated (no transgenic, knockout, or other genetically modified strains)
- The animals must be on an approved IACUC protocol for use in research or teaching
- Have not experienced any manipulation that permanently altered their anatomical, physiological, metabolic, or locomotor function
    - Post-mortem examination in which the abdomen and/or thoracic cavity has been opened or in which reproductive organs have been removed post-mortem are acceptable for donation
- Have not received any type of device implantation
- Have not received any chemical or drug, excluding isoflurane anesthesia when used in recovery procedures
- Appear free of any detectable infectious or physical abnormality that would threaten animal or public health
- Have been euthanized only through the use of CO_2 or cervical dislocation alone
- Ear tags must be removed prior to donation

The accepting entity acknowledges and has accepted the following, signed by a waiver of release.

- That the animals have been a subject in research, teaching, or testing
- Any and all risks of accepting this animal into their premises
- That the animal carcasses are being donated as a food source for animals and that the carcasses will not be sold or given away or otherwise released from his/her care
- To be responsible for all transportation of rodents by coordinating pickup with the Purdue Laboratory Animal Program

Approved by IACUC 02/16/2022

PURDUE UNIVERSITY | Office of Research
Laboratory Animal Program, Training Coordinator

Annual Isoflurane Vaporizer service dates for Spring 2024!

The Office of Research Laboratory Animal Program will be the funding the price of a **trip charge** for all on-campus research Isoflurane vaporizers for annual maintenance scheduled **May 7, 8, & 9, 2024**. (Certain buildings use the servicing company for a whole day. MJIS and Vet Med complex will have their own dates as they have many machines to service.) Our vendor, Vetamac, has arranged to be on campus to service machines over the dates listed.

Trip Charge = cost to visit Purdue	**Service Charge** = Cost to inspect the equipment

The Service Charge, the cost of any needed maintenance, repair, and/or replacement parts will be invoiced to the department owning the machine.

The IACUC requires an annual service date on all "traditional" vaporizers. We fully understand that for some, this means your service will come early, and others, the service date will be late. We have communicated to the IACUC that this will be the case for the first year of implementation. Documentation of most recent service will need to be provided in the event of an IACUC inspection.

If your equipment was serviced during our December event, please do NOT register for the May event.
Registration for the May event will open in February.
To Register for the December event, use the Office Forms link below. Please provide the following information:

- PI name and Contact Information
- Lab Manager name and Contact Information
- Business Office Contact Information
- Vaporizer Model Number
- Vaporizer Serial Number
- Equipment Location; (building, room number).

Registration ends on **April 12, 2024**. After registration you will receive a printable service tag and payment tag to attach to the equipment to be serviced along with a schedule with the date for service. After the equipment has been serviced, the technician will leave a summary tag of the service. Additionally, if a machine is no longer used, it will need to be clearly marked with a tag that states Machine NOT IN USE and it will be required to be serviced before it can be used again.

To Register, please fill out this form: https://forms.office.com/r/fQqa9k845c

Questions? Please contact Carol Dowell at dowellc@purdue.edu

VETAMAC ANESTHESIA SERVICE & SALES

Vetamac is always on campus every **3rd Monday of the month** to help you with your needs outside of these scheduled dates.

Visit: https://www.vetamac.com/ **If service by Vetamac is requested outside the service event dates, all charges and repairs, including the trip charge, will be the machine owner's responsibility.**

Registration will close on Friday, April 12, 2024 at 3:00 PM.

Vetamac Representative
Heidi Lescun, BS, RVT
Director of Technical Services
Mobile 765-479-1767
heidi.lescun@vetamac.com
130 Roth Court, Suite 100
PO Box 178
Rossville, IN 46065
(800)334-1583
(765)379-9722 Fax

Thank you,
Carol Dowell
Office of Research LAP, Training Coordinator

915 Mitch Daniels Blvd, Lilly Bldg. • West Lafayette, IN 47907
(765) 494-2521 • E-mail: dowellc@purdue.edu

Spring 2024 Rodent Tattooing for Identification!

I will be offering tattoo workshops with a maximum of 4 participants per session. (If you have any questions or special requests, please do not hesitate to contact me; 765-494-2521or dowellc@purdue.edu)

These hands-on workshops are designed to introduce the participant to the basic techniques of identification by tattoo, in the laboratory rat and mouse. A minimum of 3 days' notice is requested for general cancellations. Please follow COVID-19 safety guidelines and cancel your appointment if you are feeling ill, or have had a potential exposure.

Workshops are filled on a first-come, first-serve basis! Workshops are free to personnel tied to Purdue protocols, Purdue staff/faculty. You are encouraged to participate in workshops that are **directly related to techniques in your protocol**. ***Participants must be familiar with their project/protocol and techniques required of their protocol prior to attending class!***

REGISTRATION:

- Location for the following workshops – Meet in Lily Hall room 3-114.

Please visit, https://www.purdue.edu/research/regulatory-affairs/animal-research/workshops-events.php and select your desired date/time, and register through the "*Sign Up*" link.

1. Tattooing for rodent identification. (Limited to 4 participants each session)
   - 3/28/24 – Thursday 8:30 – 10:15am
   - 4/04/24 – Thursday 1:00 – 2:45pm
   - 5/06/24 – Monday 1:00 – 2:45pm

*We will discuss various methods for individual identification in rodents, with a focus on tattoo.

Tattooing for permanent animal identification in research animals

Purdue University Laboratory Animal Program is now offering classes for tattooing as a method of permanent animal identification.

You may join classes presented by our Training Coordinator to learn these techniques, or you can hire LAP technicians to tattoo your animals for you. (By appointment: $75/hour, minimum 30 minutes)

COMING SOON - LAP tattooing equipment is available for rent! **For more information on classes, renting or hiring technical help, please contact ddclap@purdue.edu**.

Basic tattooing can be accomplished by creating a series of dots or lines in any color, or a standard tattooing machine can be used to create numbers, lines, etc.

1. Animal tattooing technique(s) must be listed on your approved animal use protocol.
2. Personnel must be trained in techniques listed on the protocol.
3. Personnel conducting the tattooing must be listed on the approved protocol.

Suggested statements to be added to your protocol:

If you plan to hire LAP to tattoo your animals

- *Animals will be individually identified using tattoo. The Purdue LAP trained staff will be hired to use the tattoo system to place numbers or lines on the base of the tail. Numbers will be limited to 3 digits per tail or ½ the length of tail.*
- *Animals will be placed in a restraint device, tail firmly held to solid surface below. Liberal amount of tissue oil will be applied, tattoo applied, tissue blotted, then animal will be returned to normal caging. Animal identification will be listed on the cage card.*

If you plan to do your own tattooing

Using Animal Lancet or Micro-Tattooing System

- *Animals will be individually identified using tattoo. Animals are restrained by hand, placed in a restraint device, or lightly anesthetized using Isoflurane Gas Anesthesia. Area wiped using gauze and 70% Alcohol, then allowed to dry. Lancet tip or Micro-Tattooing needle is dipped into the ink, then used to make 3 puncture holes per site. A gauze pad is used to blot the excess ink from the animal. Animal returned to normal caging. (This technique is simply used to create visual dots for identification.)*

Using tattoo equipment

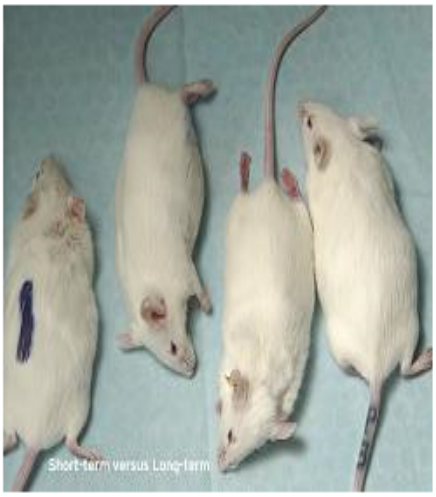
- *Prior to individually identifying animals by tattoo, participants will have documented hands-on training by with the LAP Training Coordinator, or other on-site professional instruction.*
- *Animals will be individually identified using tattoo equipment. Animals will be placed in an appropriate restraint device or lightly anesthetized. The tail will be firmly held to solid surface below. Liberal mount of tissue oil applied, tattoo marking applied, tissue blotted and inspected, then animal will be returned to normal caging. Animal identification listed on the cage card.*

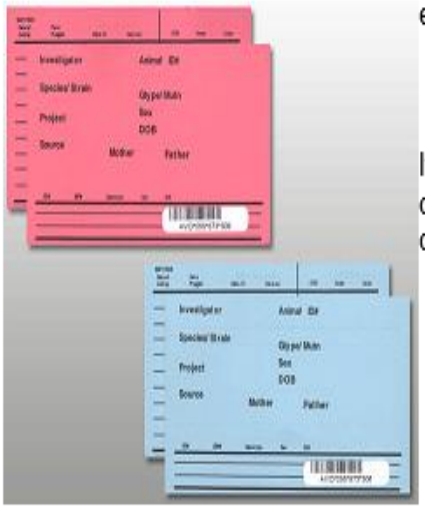
Identification Methods

It is important to select the appropriate identification method for your research purposes.

The method of identification selected must be described in the Animal Use Protocol.

Your choice of identification should be based on the age of the animal you wish to identify, the number of characters you wish to include and the duration of your experiment.





It is recommended that you record the identification information on the cage card in the event that clarification of the numbers or characters becomes necessary for any reason.

Indelible markers can be used for short-term identification. Fur clipping or shaving would also be an example of temporary marking.

Alternatively, ear punches, microchips, and tattooing are all permanent procedures.

Ear tags can be long-term, but there is always a chance they can become detached from the ear.

Consult the LAP veterinary staff if you have questions on selecting an appropriate identification method for your animals. 765-496-1019.

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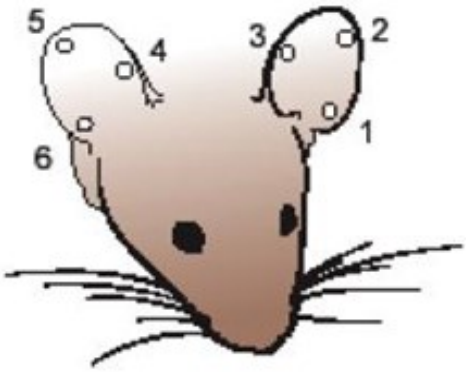
Temporary Identification

Non-toxic, permanent markers can be used to temporarily mark the fur, tail or skin of the animal.

This ink, depending on the location, usually lasts 3 - 4 days without the need to remark.

Ear Punches



Different types of ear punches are available.

Ear punches should be sterile prior to initial use.

Extra ear punches should always be available, as they become dull with repeated use.

To identify the mouse by ear punch, restrain it by the scruff using one of the methods demonstrated in the restraint section of this program.

The punch should be placed approximately 3 mm from the edge of the ear pinna.

If the punch is placed too close to the edge of the pinna, it is likely to tear and become difficult to read.

You should also be careful not to place the punch too far towards the inside of the ear to avoid injuring the animal.

The tissue removed with the ear punch can be used for genotyping.

Sanitize the ear punch between each cage of animals with 70% ethanol.

Using a chlorinated compound will cause the punch to become corroded.

Re-sterilize the instrument after use.

Microchip Transponders

The microchip transponders are implanted subcutaneously between the scapulae for permanent identification of individual animals.

Each microchip is encrypted with a unique, non-replicable number.

The chips are read with a portable, hand-held scanner. Some implants can provide you with the animal body temperature as well as identification.

To implant these chips, the mouse must be briefly anesthetized.

The hair is removed from the insertion site by shaving or plucking.

The area is prepped with an iodophor or antimicrobial skin cleanser, followed by alcohol.

The implantation needle, with the syringe attached, is purchased in a sterile package.

Make a tent from the loose skin at the implant site.

Insert the needle subcutaneously, with the bevel up, and depress the plunger.

Once the needle is removed, the injection site should be observed for bleeding.

If bleeding is noted, digital pressure with a sterile gauze pad should be applied.

If necessary, a drop of surgical glue can be applied to the needle entry site.

Tattooing

Tattooing can be used on both neonates and adults as a permanent method of identification.

Anesthesia is not required, but can be used, if necessary, to immobilize the animal.

LAP owns the AIMS System, which consists of a tattoo machine that can be used to write numbers or other characters on the tails of adult mice.

It can also be used to tattoo the footpads of both neonatal and adult mice.

The use of tattoo equipment requires training beyond this article. You may contact Carol Dowell dowellc@purdue.edu for further information. She will also have several workshops later this semester.

AIMS provides a course and certification program.

Consult your Veterinarian, Training Coordinator or equipment manufacturer for further instructions on performing tattooing.

One example of a numbering scheme is shown below.

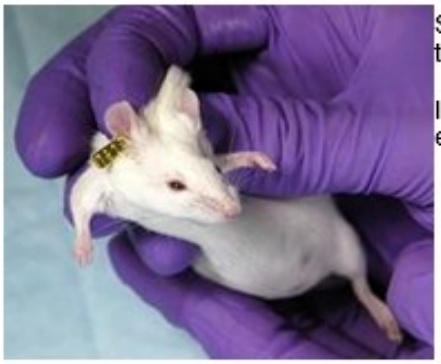
It is important to prevent potential cross contamination associated with the use of this equipment.

The needles should be either sterilized or discarded after each use.

Ear Tags

Ear tags are another means for identifying mice.

Ear tags can be imprinted by the manufacturer with several digits or letters.



Special attention should be given to the proper placement of the ear tag.

Improperly placed ear tags can become easily detached from the ear.

The tags can also be torn out when the mice fight or may inadvertently become caught in the wire bar lid.

Restrain the mouse by the one or two-hand method.

Place a sterile ear tag into a sanitized ear tag applier.

Locate the proper position for placement.

For example, place the numbers in the upward configuration so that they can be easily read without restraining the animal.

Apply the tag to the base of the ear, approximately 3 mm from the edge of the ear pinna.

Do not apply the tag too close to the center of the ear. This may cause excessive inflammation, necessitating removal of the tag.

RapID Tag: A Mouse-Eye View

Do not apply the tag too close to the outer edge of the pinna.

This may cause the tag to become entangled with the foot of the animal or in the wire bar lid, causing it to become detached from the ear.

Remember

- Select the most appropriate type of identification for your research purposes.
- The method of identification must be described in the Animal Use Protocol.
- Proper placement is essential for the use of ear punches or ear tags.
- Identification procedures should be initiated with sterile instruments. In addition, these instruments should be sanitized between cages of animals.
- Consult your veterinarian if you have questions on the selection or use of identification methods.

Source: US National Institutes of Health

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